Novel 5-Desmethylene Analogues of 5,10-Dideaza-5,6,7,8-tetrahydrofolic Acid as Potential Anticancer Agents

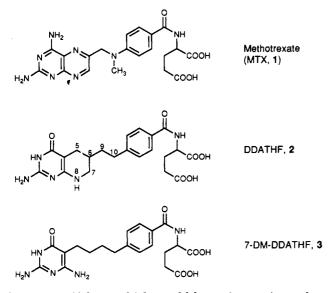
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The synthesis and biological activity of novel 5-desmethylene analogues of 5,10-dideaza-5,6,7,8-tetrahydrofolic acid (DDATHF), a potent antitumor agent presently undergoing clinical trials, are described. These compounds are representative of a new series of optically pure analogues of DDATHF.

In the treatment of cancer, one of the primary programs available to the clinician is chemotherapy. Methotrexate (MTX, 1), which represents a class of chemotherapeutic agents known as folate antimetabolites, was first prepared in 1947¹ and has in the intervening years has been the most widely used chemotherapeutic agent either alone or in combination therapy in the treatment of a wide variety of tumors.² MTX exerts its cytostatic effects in large part by inhibition of dihydrofolate reductase (DHFR), an essential enzyme in the biosynthesis of purines and pyrimidines and therefore DNA.³ However, severe side effects are normally associated with MTX therapy, and as a consequence an intensive search has been underway for more selective and less toxic antitumor agents. 5,10-Dideaza-5,6,7,8-tetrahydrofolic acid (DDATHF, Lometrexol, 2) was first described by us in 1985 and came from a search



for new antifolates which would be active at sites other than DHFR.⁴ DDATHF exhibits its cytostatic effects by inhibition of glycinamide ribonucleotide formyltransferase

(GARFT), a folate-requiring enzyme which catalyses the first of two one-carbon transfers in purine biosynthesis.⁵ The resulting inhibition of de novo purine biosynthesis provides an effective and novel mechanism of cytotostatic activity and defines a new class of antifolates with remarkable antitumor activity.⁶ Biochemical determinants of selectivity and resistance established the importance of polyglutamation for the action of DDATHF,⁷ whose transport mechanism has been shown to be the same as that of MTX.⁷ DDATHF possesses potent antitumor activity against a broad spectrum of solid tumors and is fully active against tumors which are resistant to DHFR inhibitors for reasons (such as overproduction of DHFR, or DHFR mutation to a low MTX affinity form) other than defects in active transport.⁷ Protocols utilizing the B-16 melanoma cell line have demonstrated that DDATHF completely inhibits tumor growth at 6.25 mg/kg per day for 10 days without evidence of host toxicity up to 100 mg/kg per day.⁵ DDATHF possesses structural features which allow it to act as a uniquely effective folate antimetabolite. The 2-amino-4-oxopyrimidine as well as the fully reduced pyridine ring ensure lack of inhibition of DHFR, and absence of nitrogens at the 5- and 10-positions precludes its participation as a cofactor in any one-carbon transfer reaction in folate metabolism. Retention of the *p*-benzoylglutamate moiety allows conversion to polyglutamates which are believed to be the biologically active form(s) of DDATHF. In addition, substitution of carbon for nitrogen at position 5 allows for increased chemical stability of DDATHF over tetrahydrofolic acid derivatives.

As a result of the remarkable therapeutic potential of DDATHF (now in phase II clinical trials), we have undertaken the synthesis of a broad variety of analogues in an attempt to optimize antitumor activity. Since the C-6 carbon of DDATHF is chiral, DDATHF as originally prepared (by catalytic reduction of the pyridine ring) consisted of two diastereomers, each of which exhibited slightly different intrinsic in vivo and in vitro properties. In an effort to examine the relationship between C-6 chirality and antitumor activity, we recently prepared 7-desmethylene DDATHF (7-DM-DDATHF, 3), which lacks the chiral center at C-6.⁸ This analogue showed in vitro and in vivo activity comparable to that of DDATHF

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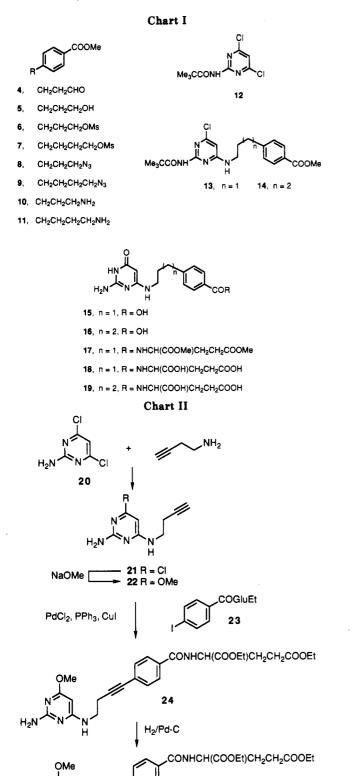
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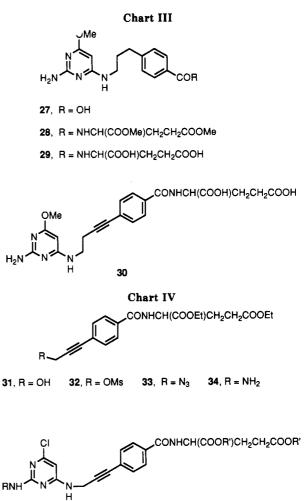
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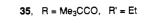
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36. B = B' = H

been eliminated by deletion of the methylene group at position 5. Our target compounds were selected to provide us with data on a range of compounds in which (i) the length and rigidity of the tether were varied and (ii) the lactam functionality in the pyrimidine ring was replaced by imino ether or imidoyl chloride functionalities.

For our synthesis of open-chain analogues 18 and 19, we required mesylates 6 and 7 (Chart I). Methyl 4-iodobenzoate underwent palladium-catalyzed coupling with allyl alcohol⁹ to give aldehyde 4.¹⁰ Sodium borohydride reduction and reaction with methanesulfonyl chloride furnished the required mesylate 6. The synthesis of 7 has been described previously.⁸ Reaction of mesylates 6 and 7 with sodium azide followed by a Staudinger reaction¹¹ gave amines 10 and 11, which underwent coupling with 2-(pivaloylamino)-4,6-dichloropyrimidine $(12)^{12}$ to give the corresponding substituted pyrimidines 13 and 14. Hydrolysis followed by glutamate coupling gave the dimethyl L-glutamate derivatives which were hydrolyzed to provide the desired target materials.

An alternative approach to compound 19 is shown in Chart II. In this case, palladium-catalyzed coupling of acetylene 22 with diethyl N-(4-iodobenzoyl)-L-glutamate¹³

itself and in addition proved to be orally effective. Encouraged by these results, we have now prepared several DDATHF analogues in which the chiral center at C-6 has

25

CONHCH(COOR)CH2CH2COOR

TMSI

26 R = Et

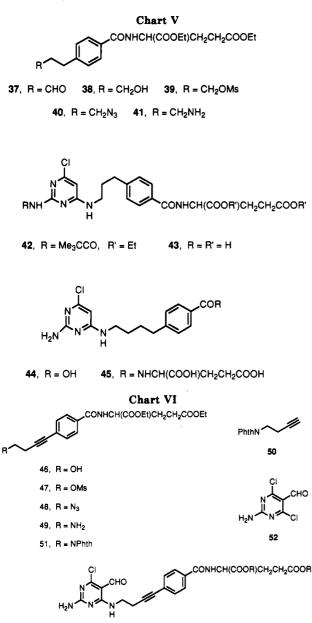
🖛 19 R = H

NaOH [

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53, R = Et 54, R = H

proceeded in poor yield, and the overall yield of 19 by this route was much lower.

In order to probe the requirement for the 4-oxo group in the pyrimidine ring, we prepared compounds 29 and 30 in which the lactam functionality is replaced by a lactim ether (Chart III). Treatment of chloropyrimidine 13 with methanolic sodium hydroxide furnished methoxypyrimidine 27 which underwent glutamate coupling and hydrolysis to furnish 29. Compound 30 was obtained by hydrolysis of diester 24.

Syntheses of analogues in which the 4-oxo functionality is replaced by a chloro substituent are outlined in Charts IV and V. Glutamate 23 underwent palladium-catalyzed coupling with propargyl alcohol to give alcohol 31. Mesylation, reaction with sodium azide, and a Staudinger reaction provided amine 34 which was condensed with 12 to give chloropyrimidine 35. Basic hydrolysis furnished target compound 36. Preparation of analogue 43 with a saturated tether necessitated the synthesis of alcohol 38. This was accomplished conveniently in two steps: palladium-catalyzed coupling of glutamate 23 with allyl alcohol gave aldehyde 37 which was reduced to the alcohol with sodium borohydride. Alcohol 38 was then converted to

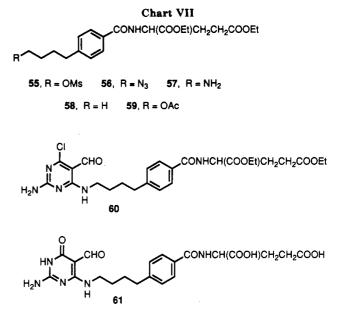


Table I. Inhibition of L1210 Cell Growth in Culture^a

compd	$IC_{50} (\mu g/mL)$	compd	$IC_{50} (\mu g/mL)$
 3	0.132	36	>20
18	0.9	43	>20
19	2.3	45	>20
29	>20	54	17.4
30	>20	61	>20

^a Data due to Dr. G. B. Grindey, Eli Lilly & Co.

target compound 43 by the sequence of reactions described above for the unsaturated alcohol 31. We had already in ester 14 an advanced intermediate in hand for the preparation of our other chloropyrimidine target 45. Acidic hydrolysis of benzoate 14 to benzoic acid 44 followed by glutamate coupling and ester hydrolysis furnished 45 in good yield.

The effect of formyl substitution in the pyrimidine ring was probed through compounds 54 and 61 (Charts VI and VII). Amine 49 was synthesized by two different methods. The first route started from alcohol 46¹⁴ and followed the same course as the reactions already described. The second route utilized a palladium-catalyzed C-C coupling reaction of glutamate 23 with acetylene 50¹⁵ followed by cleavage of the phthalimide protecting group in the resulting coupled product by treatment with methylamine¹⁶ to give the unsaturated amine 49. The corresponding saturated amine (57) was obtained by a Staudinger reaction on azide 56 which was prepared in the usual manner. We had initially planned to synthesize amine 57 by catalytic hydrogenation of the unsaturated azide 48. However, we found it impossible to obtain good yields of 57 by this approach: in alcoholic solvents, low yields of 57 were obtained, whereas in acetic acid, the unexpected products 58 and 59 were isolated. Amines 49 and 57 underwent coupling with 2-amino-4,6-dichloro-5-formylpyrimidine (52)¹⁷ to give esters 53 and 60 which were subjected to alkaline hydrolysis to provide compounds 54 and 61.

Biological Activity. The above new folate analogues were examined for their ability to inhibit the growth of L1210 cells in culture. The results of these studies are

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shown in Table I; none of these new analogues exhibited significant cytostatic activity.

Experimental Section

Methyl 4-(2-Formylethyl)benzoate (4). A mixture of methyl 4-iodobenzoate (4 g, 15.27 mmol), allyl alcohol (1.56 mL, 22.90 mmol), NaHCO₃ (3.21 g, 38.18 mmol), Pd(OAc)₂ (103 mg, 0.46 mmol), (n-Bu)₄NBr (4.92 g, 15.27 mmol), and 4-Å sieves (3 g) in DMF (50 mL) was stirred at 25 °C for 72 h. The reaction mixture was filtered through Celite and the filtrate poured onto water and extracted with ether (7 × 100 mL). The combined extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo. Flash chromatography¹⁸ (silica gel, 30% ether-hexanes) gave 2.67 g (91%) of 4 as a clear oil: ¹H NMR (CDCl₃, 300 MHz) & 2.78 (t, 2 H, J = 7 Hz), 2.96 (t, 2 H, J = 7 Hz), 3.88 (s, 3 H), 7.25 and 7.94 (AA'BB', 4 H), 9.79 (s, 1 H); EIMS m/e 192 (M⁺), 161 (base), 150, 133, 105, 91, 77; HRMS calcd for C₁₁H₁₂O₃ 192.0783, found 192.0802.

Methyl 4-(3-Hydroxypropyl)benzoate (5). Sodium borohydride (1.51 g, 39.69 mmol) was added in portions to a solution of aldehyde 4 (5.08 g, 26.46 mmol) in CH₃OH (100 mL) at 0 °C, and the resulting solution was stirred at 25 °C for 1 h. The reaction mixture was poured onto water and extracted with EtOAc $(4 \times 100 \text{ mL})$. The combined extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo. Flash chromatography (silica gel, 50% ether-hexanes) gave 4.94 g (96%) of 5 as a clear oil: ¹H NMR (CDCl₃, 300 MHz) δ 1.91 (m, 2 H, J = 7 Hz), 2.77 (t, 2 H, J = 7 Hz), 3.68 (t, 2 H, J = 7 Hz), 3.90 (s, 3 H), 7.27 and 7.96 (AA'BB', 4 H); EIMS m/e 194 (M⁺), 176, 145 (base), 117, 91, 77; HRMS calcd for C₁₁H₁₄O₃ 194.0939, found 194.0962. Anal. Calcd for C₁₁H₁₄O₈: C, 68.02; H, 7.27. Found: C, 67.98; H, 7.46.

Methyl 4-[3-(Methanesulfonyloxy)propyl]benzoate (6). To a solution of alcohol 5 (1.34 g, 6.91 mmol) in THF (30 mL) at 0 °C was added TEA (1.45 mL, 10.37 mmol) followed by MsCl (0.63 mL, 8.29 mmol), and the resulting solution was allowed to stir at 25 °C for 3 h. The reaction mixture was poured onto water and extracted with CH_2Cl_2 (3 × 100 mL). The combined extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo. Flash chromatography (silica gel, 50% ether-hexanes) gave 1.85 g (97%) of 6 as a pale yellow solid: mp 42-45 °C; ¹H NMR (CDCl₃, 300 MHz) δ 2.11 (m, 2 H, J = 7 Hz), 2.82 (t, 2 H, J = 7 Hz), 3.00 (s, 3 H), 3.90 (s, 3 H), 4.23 (t, 2 H, J = 7 Hz), 7.26 and 7.96 (AA'BB', 4 H); EIMS m/e 272 (M⁺), 212, 176, 145 (base), 115, 91; HRMS calcd for $C_{12}H_{16}O_5S$ 272.0714, found 272.0725. Anal. Calcd for $C_{12}H_{16}O_5S$: C, 52.93; H, 5.92. Found: C, 52.74; H, 6.10.

Methyl 4-(3-Azidopropyl)benzoate (8). Sodium azide (5.07 g, 77.79 mmol) was added to a solution of mesylate 6 (7.07 g, 25.99 mmol) in DMF (50 mL) at 25 °C, and the resulting solution was stirred at 25 °C for 18 h. The reaction mixture was poured onto water and extracted with CH_2Cl_2 (2 × 100 mL). The combined extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo. Flash chromatography (silica gel, 50% ether-hexanes) gave 5.28 g (93%) of 8 as a pale yellow oil: ¹H NMR (CDCl₃, 300 MHz) δ 1.92 (m, 2 H, J = 7 Hz), 2.76 (t, 2 H, J = 7 Hz), 3.29 (t, 2 H, J = 7 Hz), 3.90 (s, 3 H), 7.25 and 7.97 (AA'BB', 4 H). Anal. Calcd for $C_{11}H_{13}N_3O_2$: C, 60.26; H, 5.98; N, 19.17. Found: C, 60.01; H, 5.89; N, 19.09.

Methyl 4-(4-Azidobutyl)benzoate (9). Azide 9 was prepared in 85% yield from mesylate 7⁸ by the procedure described for the preparation of 8: ¹H NMR (CDCl₃, 270 MHz) δ 1.57–1.74 (m, 4 H), 2.68 (t, 2 H, J = 6.9 Hz), 3.26 (t, 2 H, J = 6.6 Hz), 3.88 (s, 3 H), 7.23 and 7.96 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 27.9, 28.2, 35.2, 51.0, 51.7, 127.9, 128.2, 129.6, 147.2, 166.8; EIMS m/e 204 (M⁺ – N₂ – H), 176, 172 (base), 162, 149, 131; HRMS calcd for C₁₂H₁₄NO₂ (M⁺ – N₂ – H) 204.1025, found 204.1022. Anal. Calcd for C₁₂H₁₅N₃O₂: C, 61.79; H, 6.48; N, 18.01. Found: C, 61.64; H, 6.50; N, 17.74.

Methyl 4-(3-Aminopropyl)benzoate (10). To a solution of azide 8 (2.06 g, 9.41 mmol) in THF (18 mL) at 25 °C was added water (0.25 mL, 14.12 mmol) and Ph₃P (2.47 g, 9.41 mmol), and the resulting reaction mixture was stirred at 25 °C for 18 h. The reaction mixture was poured onto 10% HCl and extracted with

ether (3 × 100 mL). The organic layer was discarded. The aqueous layer was made basic (pH = 9) with 10% NaOH and extacted with CH₂Cl₂ (4 × 100 mL). The combined extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo to give 1.78 g (98%) of 10 as a pale yellow oil: ¹H NMR (CDCl₃, 300 MHz) δ 1.20 (br s, 1 H), 1.73 (m, 2 H, J = 7 Hz), 2.70 (app q, 4 H), 3.87 (s, 3 H) 7.23 and 7.93 (AA'BB', 4 H); EIMS m/e 193 (M⁺), 176 (base), 161, 148, 116, 91, 77; HRMS calcd for C₁₁H₁₅NO₂ 193.1103, found 193.1110.

Methyl 4-(4-Aminobutyl)benzoate (11). Amine 11 was prepared in 95% yield from azide 9 by the procedure described for the preparation of 10: mp 120–122 °C; ¹H NMR (CDCl₃, 300 MHz) δ 1.42–1.47 (m, 2 H), 1.50–1.63 (m, 2 H), 1.66 (br s, 2 H), 2.50–2.69 (m, 4 H), 3.85 (s, 3 H), 7.20 and 7.92 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 75.6 MHz) δ 27.8, 32.6, 35.2, 41.4, 51.4, 127.2, 127.9, 129.1, 147.5, 166.5; EIMS m/e 207 (M⁺), 192, 172, 162, 150 (base), 131; HRMS calcd for C₁₂H₁₇NO₂ 207.1259, found 207.1258.

Methyl 4-[3-[[6-Chloro-2-(pivaloylamino)pyrimidin-4yl]amino]propyl]benzoate (13). Triethylamine (1.4 mL, 10.16 mmol) was added to a solution of amine 10 (1.78 g, 9.24 mmol) and pyrimidine 12^{12} (2.74 g, 11.09 mmol) in THF (20 mL) at 25 °C, and the resulting reaction mixture was stirred at 25 °C for 2 h. The reaction mixture was poured onto water and extracted with CH₂Cl₂ (3 × 100 mL). The combined extracts were dried over anhydrous Na₂SO₄ and concentrated in vacuo. Flash chromatography (silica gel, 50% ether-hexanes) gave 3.46 g (93%) of 13 as a white foam: ¹H NMR (CDCl₃, 300 MHz) δ 1.26 (s, 9 H), 1.94 (m, 2 H), 2.7 (m, 4 H), 3.88 (s, 3 H), 6.00 (s, 1 H), 7.20 and 7.93 (AA'BB', 4 H), 7.82 (s, 1 H); EIMS m/e 404 (M⁺), 284, 256, 242 (base), 213, 171, 158, 145, 97, 85; HRMS calcd for C₂₀-H₂₅ClN₄O₃ 404.1615, found 404.1601.

Methyl 4-[3-[[6-Chloro-2-(pivaloylamino)pyrimidin-4yl]amino]butyl]benzoate (14). Ester 14 was prepared in 61% yield from amine 11 by the procedure described for the preparation of 13: ¹H NMR (CDCl₃, 300 MHz) δ 1.29 (s, 9 H), 1.50–1.80 (m, 4 H), 2.68 (t, 2 H, J = 7.2 Hz), 3.91 (s, 3 H), 6.15 (br s, 2 H), 7.22 and 7.95 (AA'BB', 5 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 20.6, 26.9, 27.7, 28.3, 35.0, 39.8, 51.5, 127.5, 128.0, 129.3, 147.1, 156.5, 163.9, 166.6, 170.6, 175.3; EIMS m/e 418 (M⁺), 329, 269, 255 (base) 228; HRMS calcd for C₂₁H₂₇ClN₄O₃ 418.1772, found 418.1761.

4-[3-[(2-Amino-1,6-dihydro-6-oxopyrimidin-4-yl)amino]propyl]benzoic Acid (15). A slurry of ester 13 (450 mg, 1.1 mmol) in 3 N NaOH (5 mL) was allowed to reflux for 20 h. The reaction mixture was cooled, acidified with excess glacial HOAc and filtered. The residue was washed with ether (30 mL) followed by water (50 mL) and dried in vacuo to provide 317 mg (95%) of 15 as a white solid: mp >300 °C; ¹H NMR (DMSO-d₆, 300 MHz) δ 1.95 (m, 2 H), 2.3 (m, 2 H), 3.2 (m, 2 H), 5.65 (s, 1 H), 6.10 (br s, 1 H) 6.40 (br s, 2 H) 7.30 and 7.85 (AA'BB', 4 H), 9.65 (br s, 1 H), 12.65 (br s, 1 H); LRFABMS m/e 289 (MH⁺), 177, 155, 119 (base), 103; HRFABMS calcd for C₁₄H₁₇N₄O₃ 289.1301 (MH⁺), found 289.1309.

4-[3-[(2-Amino-1,6-dihydro-6-oxopyrimidin-4-yl)amino]butyl]benzoic Acid (16). Acid 16 was prepared as a white solid in 64% yield from ester 14 by the procedure described above for the preparation of 15: mp >300 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 1.44–1.57 (m, 4 H), 2.63 (br s, 2 H), 3.03 (br s, 2 H), 4.40 (s, 1 H), 6.20 (br s, 2 H), 6.35 (br s, 1 H), 7.29 and 7.83 (AA'BB', 4 H), 9.84 (br s, 1 H), 12.60 (br s, 1 H); HRFABMS calcd for $C_{15}H_{19}N_4O_3$ (MH⁺) 303.1457, found 303.1440.

Dimethyl N-[4-[3-[(2-Amino-1,6-dihydro-6-oxopyrimidin-4-yl)amino]propyl]benzoyl]-L-glutamate (17). To a stirred slurry of acid 15 (300 mg, 1.04 mmol) in DMF (4 mL) at 25 °C was added N-nethylmorpholine (NMM) (0.14 mL, 1.25 mmol) followed by 2-chloro-4,6-dimethoxy-1,3,5-triazine¹⁹ (201 mg, 1.15 mmol), and the resulting reaction mixture was stirred at 25 °C for 40 min. NMM (0.14 mL, 1.25 mmol) was added to the solution followed by dimethyl L-glutamate hydrochloride (285 mg, 1.35 mmol), and the resulting reaction mixture was allowed to stir at 25 °C for 24 h. The reaction mixture was concentrated in vacuo and the residue taken up CHCl₃ (50 mL). The CHCl₃ layer was washed with 5% NaHCO₃, dried over anhydrous Na₂SO₄, and

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concentrated in vacuo. Flash chromatography (silica gel, 7% $CH_3OH-CH_2Cl_2$) gave 256 mg (55%) of 17 as a clear foam: ¹H NMR ($CDCl_3$, 300 MHz) δ 1.85 (m, 2 H) 2.1 (m, 1 H), 2.15 (m, 1 H), 2.45 (m, 2 H), 2.60 (m, 2 H), 3.0 (m, 2 H), 3.6 (s, 3 H), 3.75 (s, 3 H), 4.75 (br s, 1 H), 4.80 (m, 1 H), 5.3 (s, 1 H), 5.5 (br s, 1 H), 6.2 (br s, 1 H), 7.05 and 7.70 (AA'BB', 4 H), 7.6 (d, 1 H, J = 7 Hz); EIMS m/e 445 (M⁺), 353, 285, 207, 167, 154 (base) 140, 111, 98, 84, 69; HRMS calcd for $C_{21}H_{27}N_5O_6$ 445.1961, found 445.1970.

N-[4-[3-[(2-Amino-1,6-dihydro-6-oxopyrimidin-4-yl)amino]propyl]benzoyl]-L-glutamic Acid (18). A solution of glutamate 17 (106 mg, 0.24 mmol) in 1 N NaOH (2.5 mL) was allowed to stir at 25 °C for 24 h. The reaction mixture was acidified with excess glacial HOAc and filtered. The residue was washed with ether (20 mL) followed by water (100 mL) and dried in vacuo to provide 82 mg (82%) of 18 as a white solid: mp 205-208 °C; ¹H NMR (DMSO-d₆, 300 MHz) δ 1.75 (m, 2 H), 1.95 (m, 1 H), 2.0 (m, 1 H), 2.30 (m, 2 H), 2.65 (m, 2 H), 3.00 (m, 2 H), 4.40 (m, 1 H), 4.40 (s, 1 H), 6.10 (br s, 2 H), 6.40 (br s, 1 H), 7.30 and 7.80 (AA'BB', 4 H), 8.50 (d, 1 H, J = 7 Hz), 9.65 (br s, 1 H), 12.40 (br s, 2 H); HRFABMS calcd for C₁₉H₂₄N₅O₆ (MH⁺) 418.1727, found 418.1744.

N-[4-[4-[(2-Amino-1,6-dihydro-6-oxopyrimidin-4-yl)amino]buty]]benzoy]-L-glutamic Acid (19). From 16. Compound 16 was coupled with dimethyl L-glutamate by the procedure described above for the preparation of 17. The crude diester was hydrolyzed by the procedure described above for the preparation of 18 to give 19 in 32% overall yield: mp >200 °C; ¹H NMR (DMSO-d₆, 300 MHz) δ 1.40-1.61 (m, 4 H), 1.86-1.97 (m, 1 H), 1.99-2.10 (m, 1 H), 2.31 (t, 2 H, J = 7.4 Hz), 2.61 (t, 2 H, J = 7.3 Hz), 3.33 (m, 2 H), 6.09 (bs s, 2 H), 6.34 (br s, 1 H), 7.26 and 7.76 (AA'BB', 4 H), 8.48 (d, 1 H, J = 7.6 Hz), 9.84 (br s, 1 H), 12.36 (br s, 2 H); HRFABMS calcd for C₂₀H₂₆N₅O₆ (MH⁺) 432.1883, found 432.1882.

From 26. Diester 26 was hydrolyzed by the procedure described above for the preparation of 18 to give 19 in 40% yield.

2-Amino-4-(3-butynylamino)-6-chloropyrimidine (21). A solution of 20 (8.20 g, 50 mmol), 3-butynylamine (3.45 g, 50 mmol) and NEt₃ (5.06 g, 50 mmol) was heated under reflux in EtOH (100 mL) for 13 h. The reaction mixture was filtered and the filtrate concentrated in vacuo. The residue was taken up in H₂O (300 mL) and extracted with CHCl₃ (2 × 300 mL). The combined extracts were dried over anhydrous MgSO₄ and concentrated in vacuo. The residue upon recrystallization from H₂O gave 5.93 g (60%) of 21 as white needles: mp 74-76 °C; ¹H NMR (CDCl₃, 300 MHz) δ 2.06 (t, 1 H, J = 2.5 Hz), 2.51 (dt, 2 H, J = 7.0, 2.5 Hz), 3.49 (m, 2 H), 4.93 (br s, 2 H), 5.10 (br s, 1 H), 5.84 (s, 1 H); EIMS m/e 196 (M⁺), 195, 157 (base), 105; HRMS calcd for C₈H₉ClN₄ 196.0516, found 196.0510. Anal. Calcd for C₈H₉ClN₄: C, 48.87; H, 4.61; N, 28.49; Cl, 18.03. Found: C, 49.14; H, 4.66; N, 28.21; Cl, 18.20.

2-Amino-4-(3-butynylamino)-6-methoxypyrimidine (22). NaOMe (2.50 g, 46 mmol) was added to a solution of 21 (4.07 g, 21 mmol) in MeOH (200 mL). The reaction mixture was heated at reflux for 24 h. An additional portion of NaOMe (2.60 g, 48 mmol) was added, and the solution was heated at reflux for 48 h, filtered to remove NaCl, concentrated to about 80 mL, and heated in a sealed tube at 140–146 °C for 4 h. The reaction mixture was filtered and the filtrate was concentrated in vacuo. Flash chromatography (silica gel, 2% MeOH–CH₂Cl₂) gave 2.17 g (54%) of 22 as an oil: ¹H NMR (CDCl₃, 300 MHz) δ 2.03 (t, H, J = 2.5 Hz), 2.48 (dt, 2 H, J = 6.5, 2.5 Hz), 3.40 (app q, 2 H, J = 6.5 Hz), 3.83 (s, 3 H), 4.79 (br s, 2 H), 4.95 (br s, 1 H), 5.18 (s, 1 H); EIMS m/e 192 (M⁺), 177, 153 (base), 140; HRMS calcd for C₉H₁₂N₄O 192.1011, found 192.1004.

Diethyl N-[4-[4-[(2-Amino-6-methoxypyrimidin-4-yl)amino]but-1-ynyl]benzoyl]-L-glutamate (24). A mixture of 22 (2.70 g, 14.05 mmol), 23¹³ (6.09 g, 14.05 mmol), PdCl₂ (55 mg, 0.31 mmol), CuI (120 mg, 0.63 mmol), PPh₃ (167 mg, 0.64 mol), and NEt₃ (5 mL) was heated under reflux in CH₃CN (100 mL) for 12 h. The reaction mixture was concentrated in vacuo. The residue taken up in CHCl₃ (200 mL) and washed with H₂O (3 × 200 mL). The CHCl₃ solution was dried over anhydrous MgSO₄ and concentrated in vacuo. Flash chromatography (silica gel, 7% MeOH-CH₂Cl₂) gave 2.07 g (30%) of 24 as an oil: ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7.2 Hz), 1.32 (t, 3 H, J = 7.2 Hz), 2.12–2.22 (m, 1 H), 2.30–2.42 (m, 1 H), 2.46–2.57 (m, 2 H), 2.73 (t, 2 H, J = 6.8 Hz), 3.49 (m, 2 H), 3.84 (s, 3 H), 4.11 (q, 2 H, J = 7.2 Hz), 4.26 (q, 2 H, J = 7.2 Hz), 4.76–4.82 (m, 3 H), 5.15 (br s, 1 H), 5.27 (s, 1 H), 7.14 (d, 1 H, J = 7.4 Hz), 7.46 and 7.76 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 13.9, 20.2, 26.9, 30.4, 40.3, 52.3, 53.1, 60.6, 61.5, 77.2, 81.4, 89.7, 126.8, 126.9, 131.5, 132.6, 162.4, 164.8, 166.5, 171.3, 171.9, 173.0; EIMS m/e 497 (M⁺), 452, 295, 266, 191, 153 (base); HRMS calcd for C₂₅H₃₁N₅O₆ 497.2274, found 497.2256.

Diethyl N-[4-[4-[(2-Amino-6-methoxypyrimidin-4-yl)amino]butyl]benzoyl]-L-glutamate (25). To a solution of 0.50 g (1 mmol) of 24 in CF₃COOH (50 mL) was added 10% Pd-C (200 mg). The mixture was hydrogenated on a Parr shaker for 4 h at 50 psi. The catalyst was filtered off through Celite and the filtrate concentrated in vacuo. The residue was taken up in $CHCl_3$ (70 mL) and washed with water (2 × 50 mL). The $CHCl_3$ solution was dried over anhydrous MgSO4 and concentrated in vacuo to give 340 mg (68%) of 25 as a yellow gum: ¹H NMR $(CDCl_3, 300 \text{ MHz}) \delta 1.24 (t, 3 \text{ H}, J = 7.1 \text{ Hz}), 1.32 (t, 3 \text{ H}, J =$ 7.1 Hz), 1.56-1.74 (m, 4 H), 2.11-2.21 (m, 1 H), 2.30-2.41 (m, 1 H), 2.45-2.56 (m, 2 H), 2.69 (t, 2 H, J = 7.2 Hz), 3.19 (m, 2 H), 3.83 (s, 3 H), 4.13 (q, 2 H, J = 7.1 Hz), 4.25 (q, 2 H, J = 7.1 Hz),4.73 (br s, 3 H), 4.77-4.83 (m, 1 H), 5.13 (s, 1 H), 7.08 (d, 1 H, J = 7.5 Hz), 7.24 and 7.75 (AA'BB', 4 H); EIMS m/e 501 (M⁺), 456, 299, 202, 181, 153 (base); HRMS calcd for C₂₅H₃₅N₅O₆ 501.2587, found 501.2616.

Diethyl N-[4-[4-[(2-Amino-1,6-dihydro-6-oxopyrimidin-4yl)amino]butyl]benzoyl]-L-glutamate (26). To a solution of 25 (210 mg, 0.42 mmol) in CHCl₃ (50 mL) was added dropwise Me₃SiI (0.6 mL, 4.2 mmol), and the resulting reaction mixture was heated under reflux for 2 h. The reaction mixture was quenched with MeOH (100 mL) and concentrated in vacuo. Flash chromatography (silica gel, 14% MeOH-CH₂Cl₂) gave 170 mg (83%) of 26 as an orange glass: ¹H NMR (CDCl₃, 300 MHz) δ 1.25 (t, 3 H, J = 7.2 Hz), 1.33 (t, 3 H, J = 7.2 Hz), 1.51–1.76 (m, 4 H), 2.13–2.23 (m, 1 H), 2.29–2.37 (m, 1 H), 2.39–2.57 (m, 2 H), 2.67–2.74 (m, 2 H), 3.42–3.50 (m, 2 H), 4.14 (q, 2 H, J = 7.2 Hz), 4.25 (q, 2 H, J = 7.2 Hz), 4.75–4.82 (m, 1 H), 5.13–5.17 (m, 1 H), 5.95 (br s, 2 H), 7.11 (d, 1 H, J = 7.6 Hz), 7.24 and 7.76 (AA'BB', 4 H), 11.20 (br s, 1 H); EIMS m/e 487, 285 (M⁺, base) 213, 202; HRMS calcd for C₂₄H₃₃N₅O₆ 487.2431, found 487.2439.

4-[3-[(2-Amino-6-methoxypyrimidin-4-yl)amino]propyl]benzoic Acid (27). A solution of ester 13 (1.08 g, 2.67 mmol) in 3 N NaOH (10 mL) and CH₃OH (2 mL) was heated to reflux for 20 h. The reaction mixture was cooled, acidified with excess glacial HOAc, and filtered. The residue was washed with ether (10 mL) and water (100 mL) and dried in vacuo to provide 0.70 g (87%) of 27 as a white solid: mp 194-196 °C; ¹H NMR (DMSO-d₆, 300 MHz) δ 1.75 (m, 2 H), 2.65 (t, 2 H, J = 7 Hz), 5.00 (s, 1 H), 5.89 (br s, 2 H), 6.60 (br t, 1 H), 7.30 and 7.82 (AA'BB', 4 H), 12.65 (br s, 1 H); EIMS m/e 302 (M⁺), 166, 154 (base), 138, 129, 125, 111, 97, 81, 71; HRMS calcd for C₁₅H₁₈N₄O₃ 302.1379, found 302.1394.

Dimethyl N-[4-[3-[(2-Amino-6-methoxypyrimidin-4-yl)amino]propyl]benzoyl]-L-glutamate (28). Diester 28 was prepared as a clear foam in 68% yield from acid 27 by the procedure described above for the preparation of 17: ¹H NMR (CDCl₃, 300 MHz) δ 1.98 (m, 2 H), 2.15 (m, 1 H), 2.35 (m, 1 H), 2.55 (m, 2 H), 2.65 (m, 2 H), 3.2 (m, 2 H), 3.63 s, 3 H), 3.76 (s, 3 H), 3.78 (s, 3 H), 4.8 (br s, 2 H), 4.8 (m, 1 H), 4.95 (br t, 1 H), 5.07 (s, 2 H), 7.2 and 7.65 (AA'BB', 4 H), 7.25 (br s, 1 H); EIMS m/e 459 (M⁺), 428, 400, 285, 167, 154 (base), 140, 125, 98; HRMS calcd for C₂₂H₂₉N₅O₆ 459.2118, found 459.2092.

N-[4-[3-[(2-Amino-6-methoxypyrimidin-4-yl)amino]propyl]benzoyl]-L-glutamic Acid (29). Diacid 29 was prepared in 64% yield from diester 28 by the procedure described above for the preparation of 18: mp 186–188 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 1.75 (m, 2 H), 1.95 (m, 1 H), 2.05 (m, 1 H), 2.35 (m, 2 H), 2.65 (m, 2 H), 3.15 (m, 2 H), 3.65 (s, 3 H), 4.40 (m, 1 H), 5.01 (s, 1 H), 5.87 (br s, 2 H), 6.56 (br t, 1 H), 7.29 and 7.78 (AA'BB', 4 H), 8.47 (d, 1 H, J = 7 Hz); LRFABMS m/e 432 (M⁺ + H), 330, 309, 279, 155, 119 (base), 103; HRFABMS calcd for C₂₀H₂₆N₅O₆ 432.1883 (MH⁺), found 432.1859.

N-[4-[4-[(2-Amino-6-methoxypyrimidin-4-yl)amino]but-1-ynyl]benzoyl]-L-glutamic Acid (30). Diacid 30 was prepared in 40% yield from diester 24 by the procedure described for the preparation of 18: mp 170 °C dec; ¹H NMR (DMSO- d_6 , 270 MHz) δ 2.04–2.28 (m, 2 H), 2.46–2.55 (m, 2 H), 2.76–2.90 (m, 2 H), 3.55–3.60 (m, 2 H), 3.82 (s, 3 H), 4.48–4.54 (m, 1 H), 5.26 (s, 1 H), 6.15 (br s, 1 H), 7.61 and 8.00 (AA'BB', 4 H), 8.82 (d, 1 H, J =7.6 Hz), 12.60 (br s, 2 H); HRFABMS calcd for C₂₁H₂₄N₅O₆ (MH⁺) 442.1727, found 442.1741.

Diethyl N-[4-(3-Hydroxy-1-propynyl)benzoyl]-L-glutamate (31). A mixture of 23 (10 g, 23.09 mmol), CuI (73 mg, 0.38 mmol), PPh₃ (100 mg, 0.38 mmol), PdCl₂ (33 mg, 0.19 mmol), and propargyl alcohol (2.22 mL, 38.16 mmol) in HNEt₂ (75 mL) was allowed to stir at 25 °C for 48 h. The reaction mixture was concentrated in vacuo. Flash chromatography (silica gel, ether) gave 7.64 g (92%) of 31 as a yellow oil: ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7 Hz), 1.32 (t, 3 H, J = 7 Hz), 1.80 (br s, 1 H), 2.20 (m, 1 H), 2.4 (m, 1 H), 2.55 (m, 2 H), 4.05 (q, 2 H, J = 7 Hz), 4.25 (q, 2 H, J = 7 Hz), 4.52 (d, 2 H, J = 5 Hz), 4.80 (m, 1 H), 7.20 (d, 1 H, J = 5 Hz), 7.47 and 7.79 (AA'BB', 4 H); EIMS m/e 361 (M⁺), 287, 257, 202, 158 (base), 129, 112, 84; HRMS calcd for C₁₉H₂₃NO₆ 361.1525, found 361.1554.

Diethyl N-[4-[3-(Methanesulfonyloxy)-1-propynyl]benzoyl]-L-glutamate (32). Mesylate 32 was prepared in 91% yield from alcohol 31 by the procedure described above for the preparation of 6: ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7 Hz), 1.32 (t, 3 H, J = 7 Hz), 2.20 (m, 1 H), 2.40 (m, 1 H), 2.55 (m, 2 H), 3.14 (s, 3 H), 4.05 (q, 2 H, J = 7 Hz), 4.25 (q, 2 H, J = 7 Hz), 4.80 (m, 1 H), 5.07 (s, 2 H), 7.20 (d, 1 H, J = 7 Hz), 7.47 and 7.79 (AA'BB', 4 H); EIMS m/e 439 (M⁺), 394, 366, 344, 237 (base), 202, 159, 142, 112, 84; HRMS calcd for C₂₀H₂₅NO₈S 439.1301, found 439.1282.

Diethyl N-[4-(3-Azido-1-propynyl)benzoyl]-L-glutamate (33). Azide 33 was prepared in 80% yield from mesylate 32 by the procedure described above for the preparation of 8: ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7 Hz), 1.32 (t, 3 H, J = 7Hz), 2.20 (m, 1 H), 2.40 (m, 1 H), 2.55 (m, 2 H), 4.05 (q, 2 H, J= 7 Hz), 4.17 (s, 2 H), 4.25 (q, 2 H, J = 7 Hz), 4.80 (m, 1 H), 7.20 (d, 1 H, J = 7 Hz), 7.47 and 7.79 (AA'BB', 4 H); EIMS m/e 386 (M⁺), 358, 312, 285, 239, 202, 156 (base), 112, 84; HRMS calcd for C₁₉H₂₂N₄O₅ 386.1590, found 386.1589.

Diethyl N-[4-(3-Amino-1-propynyl)benzoyl]-L-glutamate (34). Amine 34 was prepared in 61% yield from azide 33 by the procedure described above for the preparation of 10: ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7 Hz), 1.32 (t, 3 H, J = 7Hz), 2.20 (m, 1 H), 2.40 (m, 1 H), 2.55 (m, 2 H), 3.65 (br s, 2 H), 4.05 (q, 2 H, J = 7 Hz), 4.25 (q, 2 H, J = 7 Hz), 4.80 (m, 1 H), 5.15 (s, 2 H), 7.30 (d, 1 H, J = 7 Hz), 7.37 and 7.71 (AA'BB', 4 H); EIMS m/e 360 (M⁺), 287, 202, 158 (base), 112, 77; HRMS calcd for C₁₉H₂₄N₂O₅ 360.1685, found 360.1666.

Diethyl N-[4-[3-[[2-(Pivaloylamino)-6-chloropyrimidin-4-yl]amino]-1-propynyl]benzoyl]-L-glutamate (35). Diester 35 was prepared in 90% yield from amine 34 by the procedure described above for the preparation of 13: ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7 Hz), 1.32 (t, 3 H, J = 7 Hz), 1.35 (s, 9 H), 1.80 (s, 2 H), 2.20 (m, 1 H), 2.35 (m, 1 H), 2.55 (m, 2 H), 4.05 (q, 2 H, J = 7 Hz), 4.25 (q, 2 H, J = 7 Hz), 4.40 (br s, 1 H), 4.80 (m, 1 H), 6.25 (s, 1 H), 7.20 (d, 1 H, J = 7 Hz), 7.45 and 7.80 (AA'BB', 4 H), 7.91 (br s, 1 H); EIMS m/e 570 (M⁺ - 2H), 513, 514, 487, 485, 370, 368, 312, 310, 285, 283, 257, 255, 246, 128, 84; HRMS calcd for C₂₈H₃₅ClN₅O₆ 572.2275, found 572.2248.

N-[4-[3-[(2-Amino-6-chloropyrimidin-4-yl)amino]-1propynyl]benzoyl]-L-glutamic Acid (36). Diacid 36 was prepared in 45% yield from diester 35 by the procedure described above for the preparation of 18: mp 138-140 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 2.00 (m, 2 H), 2.2 (m, 2 H), 4.40 (m, 3 H), 5.90 (s, 1 H), 6.6 (br s, 2 H), 7.50 and 7.90 (AA'BB', 4 H), 7.60 (br s, 1 H), 8.60 (br s, 1 H), 12.50 (br s, 2 H); LRFABMS, m/e432 (M⁺), 287, 285, 229, 267, 155, 119 (base); HRFABMS calcd for C₁₉H₁₈ClN₅O₅ 432.1075, found 432.1069.

Diethyl N-[4-(2-Formylethyl)benzoyl]-L-glutamate (37). Aldehyde 37 was prepared in 74% yield from 23¹³ by the procedure described above for the preparation of 4: mp 75–76 °C; ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7 Hz), 1.28 (t, 3 H, J = 7 Hz), 2.15 (m, 1 H), 2.35 (m, 1 H), 2.55 (m, 2 H), 2.83 (t, 2 H, J = 7 Hz), 3.03 (t, 2 H, J = 7 Hz), 4.15 (q, 2 H, J = 7 Hz), 4.30 (t, 2 H, J = 7 Hz), 4.15 (q, 2 H, J = 7 Hz), 7.30 and 7.78 (AA'BB', 4 H), 9.85 (s, 1 H); EIMS m/e 363 (M⁺), 290, 202, 161 (base), 131, 112, 91, 84; HRMS calcd for C₁₉H₂₅NO₆ 363.1682, found 363.1667. Anal. Calcd for $C_{19}H_{25}NO_6$: C, 62.80; H, 6.93; N, 3.85. Found: C, 62.81; H, 6.98; N, 4.00.

Diethyl N-[4-(3-Hydroxypropyl)benzoyl]-L-glutamate (38). Alcohol 38 was prepared in 93% yield by the procedure described above for the preparation of 5: ¹H NMR (CDCl₃, 300 MHz) δ 1.20 (t, 3 H, J = 7 Hz), 1.35 (t, 3 H, J = 7 Hz), 1.95 (t, 2 H, J = 7 Hz), 2.15 (m, 1 H), 2.35 (m, 1 H), 2.55 (m, 2 H), 2.80 (t, 3 H, J = 7Hz), 3.80 (br s, 2 H), 4.05 (q, 2 H, J = 7 Hz), 4.20 (q, 2 H, J = 7Hz), 4.80 (m, 1 H), 7.05 (d, 1 H, J = 7 Hz), 7.25 and 7.74 (AA'BB', 4 H); EIMS, m/e 365 (M⁺), 292, 202, 163 (base), 145, 112, 91; HRMS calcd for C₁₉H₂₇NO₆ 365.1838, found 365.1817.

Diethyl N-[4-[3-(Methanesulfonyloxy)propyl]benzoyl]-L-glutamate (39). Mesylate 39 was prepared in 94% yield by the procedure described above for the preparation of 6: ¹H NMR (CDCl₃, 300 MHz) δ 1.20 (t, 3 H, J = 7 Hz), 1.35 (t, 3 H, J = 7Hz), 2.10 (m, 3 H), 2.35 (m, 1 H), 2.55 (m, 2 H), 2.80 (t, 2 H, J = 7 Hz), 3.02 (s, 3 H), 4.15 (m, 4 H), 4.20 (m, 2 H), 4.80 (m, 1 H), 7.10 (d, 1 H, J = 7 Hz), 7.25 and 7.80 (AA'BB', 4 H); EIMS m/e443 (M⁺), 370, 241, 202, 161, 145, 131, 112, 91; HRMS calcd for C₂₀H₂₉NO₈S 443.1614, found 443.1595.

Diethyl N-[4-(3-Azidopropyl)benzoyl]-L-glutamate (40). Azide 40 was prepared in 93% yield by the procedure described above for the preparation of 8: ¹H NMR (CDCl₃), 300 MHz), δ 1.15 (t, 3 H, J = 7 Hz), 1.25 (t, 3 H, J = 7 Hz), 1.85 (m, 2 H), 2.05 (m, 1 H), 2.15 (m, 2 H), 2.4 (m, 2 H), 2.65 (t, 2 H, J = 7 Hz), 3.20 (t, 2 H, J = 7 Hz), 4.0 (q, 2 H, J = 7 Hz), 4.15 (q, 2 H, J = 7 Hz), 4.75 (m, 1 H), 7.15 and 7.75 (AA'BB', 4 H), 7.30 (d, 1 H, J = 7Hz); EIMS m/e 390 (M⁺), 362, 335, 317, 289, 202, 188, 160 (base), 131, 112, 90; HRMS calcd for C₁₉H₂₆N₄O₅ 390.1903, found 390.1912.

Diethyl N-[4-(3-Aminopropyl)benzoyl]-L-glutamate (41). Amine 41 was prepared in 95% yield by the procedure described above for the preparation of 10: ¹H NMR (CDCl₃, 300 MHz) δ 1.20 (t, 3 H, J = 7 Hz), 1.35 (t, 3 H, J = 7 Hz), 1.80 (m, 2 H), 2.10 (m, 1 H), 2.35 (m, 1 H), 2.45 (m, 2 H), 2.70 (m, 4 H), 3.2 (br s, 2 H), 4.05 (q, 2 H, J = 7 Hz), 4.25 (q, 4 H, J = 7 Hz), 4.80 (m, 1 H), 7.20 (m, 1 H), 7.25 and 7.75 (AA'BB', 4 H); EIMS m/e 364 (M⁺), 334, 291, 274, 202, 162 (base), 145, 130, 112, 91; HRMS calcd for C₁₉H₂₈N₂O₅ 364.1998, found 364.1986.

Diethyl N-[4-[3-[[2-(Pivaloylamino)-6-chloropyrimidin-4-yl]amino]propyl]benzoyl]-L-glutamate (42). Diester 42 was prepared in 82% yield by the procedure described above for the preparation of 13: ¹H NMR (CDCl₃, 300 MHz) δ 1.2 (m, 6 H), 1.31 (s, 9 H), 1.95 (m, 4 H), 2.2 (m, 1 H), 2.35 (m, 1 H), 2.55 (m, 2 H), 2.8 (m, 2 H), 3.2 (br s, 1 H), 4.15 (q, 2 H, J = 7 Hz), 4.25 (q, 2 H, J = 7 Hz), 4.80 (m, 1 H), 6.00 (s, 1 H), 7.15 (d, 1 H, J= 7 Hz), 7.25 and 7.75 (AA'BB', 4 H), 7.90 (br s, 1 H); EIMS m/e575 (M⁺), 518, 315, 242 (base), 158, 98, 84; HRMS calcd for C₂₈H₃₈ClN₅O₆ 575.2511, found 575.2524.

N-[4-[3-[(2-Amino-6-chloropyrimidin-4-yl)amino]propyl]benzoyl]-L-glutamic Acid (43). Diacid 43 was prepared in 62% yield from diester 42 by the procedure described above for the preparation of 18: mp 198-200 °C; ¹H NMR (CDCl₃, 300 MHz) δ 2.00 (m, 2 H), 2.2 (m, 1 H) 2.35 (m, 1 H), 2.55 (br s, 2 H), 2.60 (m, 2 H), 2.95 (t, 2 H, J = 7 Hz), 4.65 (m, 1 H), 6.00 (s, 1 H), 6.65 (br s, 2 H), 7.40 (br s, 1 H), 7.6 and 8.10 (AA'BB', 4 H), 8.80 (d, 1 H, J = 7 Hz), 12.65 (br s, 1 H); LRFABMS m/e436 (M⁺), 311, 309, 167, 157, 155, 135, 119 (base); HRFABMS calcd for C₁₉H₂₂ClN₅O₅ 436.1388, found 436.1384.

4-[4-[(6-Chloro-2-aminopyrimidin-4-yl)amino]buty]]benzoic Acid (44). A solution of 14 (1.20 g, 2.86 mmol) in 6 M HCl (50 mL) was heated under reflux for 3 h. Solid NaHCO₃ was added to the cooled solution to bring the pH to 5, and the precipitated solid was filtered, washed with water, and dried in vacuo to give 650 mg (71%) of 44 as a white solid: mp 133-135 °C; ¹H NMR (DMSO-d₆, 300 MHz) δ 1.44-1.65 (m, 4 H), 2.62 (t, 2 H, J = 7.3 Hz), 3.10-3.35 (br s, 1 H), 5.69 (s, 1 H), 6.37 (br s, 2 H), 7.14 (br s, 1 H), 7.28 and 7.82 (AA'BB', 4 H), 12.72 (br s, 1 H); EIMS, m/e 320 (M⁺), 185, 171, 157 (base), 144; HRMS calcd for C₁₅H₁₇ClN₄O₂ 320.1040, found 320.1041.

N-[4-[4-[(6-Chloro-2-aminopyrimidin-4-yl)amino]butyl]benzoyl]-L-glutamic Acid (45). Diacid 45 was prepared in 56% yield from 44 by the procedure described above for the preparation of 19 from 16: mp 112–114 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 1.42–1.49 (m, 2 H), 1.53–1.62 (m, 2 H), 1.87–1.97 (m, 1 H), 2.00–2.11 (m, 1 H), 2.31 (t, 2 H, J = 7.3 Hz), 2.62 (t, 2 H, J = 7.2 Hz), 3.22 (br s, 2 H), 4.31–4.38 (m, 1 H), 5.72 (s, 1 H), 6.45 (br s, 2 H), 7.25 (m, 3 H), 7.76 (d, 2 H, J = 8.0 Hz), 8.50 (d, 1 H, J = 7.7 Hz), 12.40 (br s, 2 H); HRFABMS calcd for $C_{20}H_{25}ClN_5O_5$ (MH⁺) 450.1544, found 450.1544.

Diethyl N-[4-[4-(Methanesulfonyloxy)-1-butyny]benzoy]-L-glutamate (47). Mesylate 47 was prepared in 99% yield from alcohol 46⁸ by the procedure described above for the preparation of 6: mp 82-84 °C; ¹H NMR (CDCl₃, 300 MHz), δ 1.23 (t, 3 H, J = 7.2 Hz), 1.31 (t, 3 H, J = 7.2 Hz), 2.11-2.19 (m, 1 H), 2.28-2.38 (m, 1 H), 2.44-2.52 (m, 2 H), 2.92 (t, 2 H, J = 6.7Hz), 3.09 (s, 3 H), 4.10 (q, 2 H, J = 7.2 Hz), 4.25 (q, 2 H, J = 7.2Hz), 4.41 (t, 2 H, J = 6.7 Hz), 4.74-4.80 (m, 1 H), 7.13 (d, 1 H, J = 7.3 Hz), 7.47 and 7.77 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 14.1, 20.8, 27.2, 30.5, 37.8, 52.5, 60.8, 61.8, 67.1, 82.2, 86.6, 126.5, 127.1, 131.8, 133.2, 166.3, 171.9, 173.3; EIMS m/e 453 (M⁺), 251 (base), 202, 155; HRMS calcd for C₂₁H₂₇NO₈S: 453.1457, found 453.1432. Anal. Calcd for C₂₁H₂₇NO₈S: C, 55.62; H. 6.00; N, 3.09; S, 7.07. Found: C, 55.41; H, 6.10; N, 3.28; S, 7.35.

Diethyl N-[4-(4-Azido-1-butynyl)benzoyl]-L-glutamate (48). Azide 48 was prepared in 84% yield from mesylate 47 by the procedure described above for the preparation of 8: mp 35–37 °C; ¹H NMR (CDCl₃, 300 MHz) δ 1.24 (t, 3 H, J = 7.0 Hz), 1.32 (t, 3 H, J = 7.0 Hz), 2.12–2.22 (m, 1 H), 2.30–2.39 (m, 1 H), 2.45–2.56 (m, 2 H), 2.76 (t, 2 H, J = 6.7 Hz), 3.51 (t, 2 H, J = 6.7 Hz), 4.12 (q, 2 H, J = 7.0 Hz), 4.26 (q, 2 H, J = 7.0 Hz), 4.77–4.82 (m, 1 H), 7.09 (d, 1 H, J = 7.3 Hz), 7.49 and 7.77 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 13.8, 20.4, 26.8, 30.2, 49.5, 52.2, 60.4, 61.3, 8.16, 88.3, 126.4, 126.8, 131.3, 132.7, 166.2, 171.6, 172.8; EIMS m/e 400 (M⁺), 372, 326, 202, 198, 170 (base); HRMS calcd for C₂₀H₂₄N₄O₅ : C, 59.99; H, 6.04; N, 13.99. Found: C, 60.25; H, 6.05; N, 14.09.

Diethyl N-[4-(4-Amino-1-butynyl)benzoyl]-L-glutamate (49). From 48. To a solution of 48 (1.25 g, 3.12 mmol) in CH₂Cl₂ (100 mL) was added PPh₃ (862 mg, 3.29 mmol) and the resulting reaction mixture was heated at reflux overnight. Water (50 mL) was added, and the resulting solution was stirred at 25 °C for 20 h. The CH_2Cl_2 layer was separated, washed with water (50 mL), dried over anhydrous MgSO4, and concentrated in vacuo. Flash chromatography (silica gel, CH₂Cl₂/NEt₃/MeOH [92:5:3]) gave 870 mg (74%) of 49 as an oil: ¹H NMR (CDCl₃, 300 MHz) δ 1.21 (t, 3 H, J = 7.1 Hz), 1.28 (t, 3 H, J = 7.1 Hz), 1.60 (br s, 2 H),2.10-2.20 (m, 1 H), 2.23-2.35 (m, 1 H), 2.42-2.52 (m, 2 H), 2.56 (t, 2 H, J = 6.0 Hz), 2.90 (t, 2 H, J = 6.2 Hz), 4.09 (q, 2 H, J =7.1 Hz), 4.20 (q, 2 H, J = 7.1 Hz), 4.73-4.78 (m, 1 H), 7.43 and 7.79 (AA'BB', 5 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 13.7, 24.0, 26.5, 30.1, 40.6, 52.0, 60.2, 61.0, 89.0, 90.3, 126.8, 131.1, 132.3, 166.3, 171.5, 172.6; EIMS m/e 374 (M⁺), 345, 301, 172, 142 (base); HRMS

calcd for $C_{20}H_{26}N_2O_5$ 374.1842, found 374.1834. **From 51.** To a solution of 51 (4.21 g, 8.34 mmol) in EtOH (100 mL) and CHCl₃ (100 mL) under N₂ was added 40% aqueous MeNH₂ (2.75 g, 35.42 mmol), and the resulting reaction mixture was stirred at 25 °C for 18 h and then concentrated in vacuo. Flash chromatography (silica gel, CH₂Cl₂/NEt₃/MeOH [92:5:3]) gave 2.34 g (75%) of 49.

Diethyl N-[4-(4-Phthalimidobut-1-ynyl)benzoyl]-L-glutamate (51). A mixture of 1.09 g (5.45 mmol) of 50,10 2.36 g (5.45 mmol) of 23, 60 mg (0.27 mmol) of Pd(OAc)₂, 50 mg (0.26 mmol) of CuI, 145 mg (0.55 mmol) of PPh₃, 5 mL of NEt₃, and 10 mL of CH₃CN was heated under reflux for 24 h. The solvent was removed to give a brown solid which was filtered through a short pad of silica gel (eluting with 3% $MeOH/CH_2Cl_2$) to give 2.41 g (4.78 mmol, 92%) of 51. The analytical sample was recrystallized from AcOH/H₂O: mp 145-146 °C; ¹H NMR (CDCl₃, 300 MHz) δ 1.22 (t, 3 H, J = 7.1 Hz), 1.30 (t, 3 H, J = 7.1 Hz), 2.10-2.20 (m, 1 H), 2.26–2.36 (m, 1 H), 2.38–2.54 (m, 2 H), 2.85 (t, 2 H, J = 6.9 Hz), 3.98 (t, 2 H, J = 7.0 Hz), 4.10 (q, 2 H, J = 7.1 Hz), 4.24 (q, 2 H, J = 7.1 Hz), 4.75-4.80 (m, 1 H), 7.00 (d, 1 H, J = 7.5 Hz),7.39 (d, 2 H, J = 8.0 Hz), 7.70–7.75 (m, 4 H), 7.90 (m, 2 H); EIMS m/e 504 (M⁺), 431, 302 (base), 202, 160; HRMS calcd for C₂₈- $H_{28}N_2O_7$ 504.1897, found, 504.1896. Anal. Calcd for $C_{28}H_{28}N_2O_7$: C, 66.66; N, 5.59; N, 5.55. Found: C, 66.46; H, 5.46; N, 5.55.

Diethyl N-[4-[4-](2-Amino-6-chloro-5-formylpyrimidin-4-yl)amino]-1-butynyl]benzoyl]-L-glutamate (53). A mixture of amine 49 (670 mg, 1.79 mmol), pyrimidine 52¹⁷ (343 mg, 1.79 mmol), and NEt₃ (183 mg, 1.81 mmol) was heated under reflux in MeOH (80 mL) for 3 h. The reaction mixture was concentrated in vacuo, and the residue taken up in CHCl₃ (100 mL) and washed with H_2O (2 × 50 mL). The CHCl₃ solution was dried over anhydrous MgSO₄ and concentrated in vacuo. Flash column chromatography (silica gel, 65% EtOAc-hexanes) gave 660 mg (70%) of 53 as a white powder: mp 153-155 °C; ¹H NMR (CDCl₃, 300 MHz) δ 1.23 (t, 3 H, J = 7.1 Hz), 1.31 (t, 3 H, J = 7.1 Hz), 2.13-2.18 (m, 1 H), 2.30-2.40 (m, 1 H), 2.43-2.52 (m, 2 H), 2.75 (t, 2 H, J = 6.8 Hz), 3.74 (m, 2 H), 4.11 (q, 2 H, J = 7.1 Hz), 4.25(q, 2 H, J = 7.1 Hz), 4.78 (m, 1 H), 5.50 (br s, 2 H), 7.06 (d, 1 H)J = 7.4 Hz), 7.50 and 7.76 (AA'BB', 4 H), 9.54 (br s, 1 H), 10.11 (s, 1 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 14.1, 20.1, 27.2, 30.5, 39.4, 52.5, 60.8, 61.7, 81.9, 94.1, 102.8, 126.9, 127.0, 131.8, 132.8, 162.3, 162.7, 166.4, 171.9, 173.2, 188.8; EIMS m/e 529 (M⁺), 327, 223 (base), 185, 155, 141; HRMS calcd for C₂₅H₂₈ClN₅O₆ 529.1728, found 529.1739. Anal. Calcd for C25H28ClN5O6: C, 56.66; H, 5.33; N, 13.21; Cl, 6.69. Found: C, 56.48; H, 5.20; N, 13.08; Cl, 6.97.

N-[4-[4-[(2-Amino-1,6-dihydro-5-formyl-6-oxopyrimidin-4-yl)amino]but-1-ynyl]benzoyl]-L-glutamic Acid (54). Compound 54 was prepared in 41% yield from 53 by the procedure described for the preparation of 18: mp 157-162 °C dec; ¹H NMR (DMSO- $d_{\rm g}$) δ 2.04-2.26 (m, 2 H), 2.49 (t, 2 H, J = 7.3 Hz), 2.88 (t, 2 H, J = 7.0 Hz), 3.78-3.86 (m, 2 H), 4.47-4.55 (m, 1 H), 7.66 and 8.01 (AA'BB', 4 H), 8.76 (d, 1 H, J = 7.6 Hz), 9.81 (s, 1 H), 10.07 (br s, 2 H), 10.88 (br s, 1 H), 12.50 (br s, 2 H); FABMS calcd for C₂₁H₂₂N₆O₇ (MH⁺) 456.1519, found 456.1510.

Diethyl N-[4-(4-Azidobutyl)benzoyl]-L-glutamate (56). Azide 56 was prepared from mesylate 55⁸ in 72% yield by the procedure described above for the preparation of 8: mp 54-55 °C; ¹H NMR (CDCl₃, 300 MHz) δ 1.22 (t, 3 H, J = 7.1 Hz), 1.31 (t, 3 H, J = 7.1 Hz), 1.59-1.75 (m, 4 H), 2.12-2.17 (m, 1 H), 2.28-2.40 (m, 2 H), 2.43-2.55 (m, 1 H), 2.70 (t, 2 H, J = 7.3 Hz), 3.30 (t, 2 H, J = 6.5 Hz), 4.11 (q, 2 H, J = 7.1 Hz), 4.23 (q, 2 H, J = 7.1 Hz), 4.77-4.82 (m, 1 H), 7.06 (d, 1 H, J = 7.5 Hz), 7.24 and 7.76 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 75.6 MHz) δ 13.8, 26.9, 27.8, 28.1, 30.2, 34.9, 37.3, 50.9, 52.0, 60.4, 61.3, 127.0, 128.2, 131.2, 145.7, 166.7, 171.7, 172.8; EIMS m/e 404 (M⁺), 375, 359, 331 (base), 202; HRMS calcd for C₂₀H₂₈N₄O₅ 404.2060, found 404.2057. Anal. Calcd for C₂₀H₂₈N₄O₅ (C, 59.39; H, 6.98; N, 13.85. Found: C, 59.52; H, 6.78; N, 13.60.

Diethyl N-[4-(4-Aminobutyl)benzoyl]-L-glutamate (57). Amine 57 was prepared from azide 56 in 95% yield by the procedure described above for the preparation of 10: ¹H NMR (CDCl₃, 300 MHz) δ 1.22 (t, 3 H, J = 7.1 Hz), 1.30 (t, 3 H, J = 7.1 Hz), 1.52–1.70 (m, 4 H), 2.12–2.20 (m, 1 H), 2.27–2.42 (m, 1 H), 2.43–2.55 (m, 2 H), 2.67 (t, 2 H, J = 7.3 Hz), 2.77 (m, 2 H), 3.60 (br s, 2 H), 4.11 (q, 2 H, J = 7.1 Hz), 4.23 (q, 2 H, J = 7.1 Hz), 4.75–4.80 (m, 1 H), 7.08 (d, 1 H, J = 7.6 Hz), 7.24 and 7.73 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 75.6 MHz) δ 14.1, 27.1, 28.2, 30.5, 35.3, 40.9, 46.1, 52.3, 60.8, 61.6, 127.3, 128.5, 131.2, 146.3, 167.2, 172.1, 173.2; EIMS m/e 378 (M⁺), 321, 204, 176 (base); HRMS calcd for C₂₀H₃₀N₂O₅ 378.2155, found 378.2154.

Diethyl N-[4-(4-Acetoxybutyl)benzoyl]-L-glutamate (59) and Diethyl N-(4-Butylbenzoyl)-L-glutamate (58). A solution of azide 48 (2.98 g, 7.44 mmol) in AcOH (50 mL) was hydrogenated over 300 mg of 10% Pd-C on a Parr shaker for 10 h at 50 psi. The reaction mixture was filtered through Celite and the filtrate concentrated in vacuo. Flash chromatography (silica gel, 40% EtOAc-hexanes) gave 290 mg (11%) of 58 as white needles [mp 53–55 °C; ¹H NMR (CDCl₃, 300 MHz) δ 0.95 (t, 3 H, J = 7.3 Hz), 1.24 (t, 3 H, J = 7.2 Hz), 1.30-1.40 (m, 5 H), 1.57-1.63 (m, 2 H), 2.11-2.19 (m, 1 H), 2.31-2.39 (m, 1 H), 2.44-2.54 (m, 2 H), 2.68(t, 2 H, J = 7.6 Hz), 4.12 (q, 2 H, J = 7.2 Hz), 4.26 (q, 2 H, J = 7.2 Hz)7.2 Hz), 4.81 (m, 1 H), 6.99 (d, 1 H, J = 5.3 Hz), 7.27 and 7.75 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 67.9 MHz) δ 13.8, 14.1, 14.1, 22.2, 27.5, 30.5, 33.2, 35.5, 52.3, 60.7, 61.6, 127.1, 128.6, 131.3, 147.2, 167.1, 172.0, 173.1; EIMS m/e 363 (M⁺), 318, 290, 202, 161 (base);HRMS calcd for $C_{20}H_{29}NO_5$ 363.2046, found 363.2029. Anal. Calcd for $C_{20}H_{29}NO_5$: C, 66.09; H, 8.04; N, 3.85. Found: C, 66.36; H, 7.81; N, 3.92] and 1.35 g (43%) of 59 as a liquid: ¹H NMR $(\text{CDCl}_3, 300 \text{ MHz}) \delta 1.22 \text{ (t, 3 H, } J = 7.0 \text{ Hz}), 1.30 \text{ (t, 3 H, } J = 7.0 \text{ Hz})$ 7.0 Hz), 1.60-1.80 (m, 4 H), 2.04 (s, 3 H), 2.06-2.20 (m, 1 H), 2.26-2.36 (m, 1 H), 2.38-2.57 (m, 2 H), 2.69 (t, 2 H, J = 6.7 Hz),4.05–4.15 (m, 4 H), 4.23 (q, 2 H, J = 7.2 Hz), 4.80 (m, 1 H), 7.11 (d, 1 H, J = 7.3 Hz), 7.24 and 7.75 (AA'BB', 4 H); ¹³C NMR (CDCl₃, 67.9 MHz) & 14.0, 20.8, 27.1, 27.3, 28.0, 30.4, 35.1, 52.2,

60.6, 61.5, 64.0, 127.1, 128.4, 131.2, 146.1, 166.9, 171.0, 171.9, 173.0; EIMS m/e 421 (M⁺), 348, 219 (base), 202, 159; HRMS calcd for $C_{22}H_{31}NO_7$ 421.2101, found 421.2104. Anal. Calcd for $C_{22}H_{31}NO_7$: C, 62.69; H, 7.41; N, 3.32. Found: C, 62.91; H, 7.27; N, 3.37.

Diethyl N-[4-[4-[(2-Amino-6-chloro-5-formylpyrimidin-4-yl)amino]butyl]benzoyl]-L-glutamate (60). Diester 60 was prepared in 71% yield from amine 57 by the procedure described above for the preparation of 53: mp 113–115 °C; ¹H NMR ($CDCl_3$, 300 MHz) δ 1.22 (t, 3 H, J = 7.1 Hz), 1.30 (t, 3 H, J = 7.1 Hz), 1.56-1.76 (m, 4 H), 2.08-2.21 (m, 1 H), 2.27-2.34 (m, 1 H), 2.40-2.53 (m, 2 H), 2.67 (t, 2 H, J = 6.8 Hz), 3.39–3.48 (m, 2 H), 4.11 (q, 2 H, J = 7.1 Hz, 4.23 (q, 2 H, J = 7.1 Hz), 4.76-4.83 (m, 1 H),5.80 (br s, 1 H), 7.15-7.23 (m, 3 H), 7.73 (d, 2 H, J = 8 Hz), 9.23 $(t, 1 H, J = 5.5 Hz), 10.04 (s, 1 H); {}^{13}C NMR (CDCl_3, 75.6 MHz)$ δ 14.0, 27.1, 28.0, 28.3, 30.4, 35.1, 40.1, 52.2, 60.6, 61.5, 102.2, 127.1, 128.4, 131.3, 146.0, 162.1, 162.3, 166.3, 167.1, 172.1, 173.0, 188.5; EIMS m/e 533 (M⁺), 505, 460, 331 (base), 302, 185; HRMS calcd for C25H32ClN5O6 533.2041, found 533.2017. Anal. Calcd for C₂₅H₃₂ClN₅O₆: C, 56.23; H, 6.04; N, 13.11; Cl, 6.64. Found: C, 56.03; H, 5.79; N, 12.88; Cl, 6.77.

N-[4-[4-[(2-Amino-1,6-dihydro-5-formyl-6-oxopyrimidin-4-yl)amino]butyl]benzoyl]-L-glutamic Acid (61). Diacid 61 was prepared in 25% yield from diester 60 by the procedure described above for the preparation of 18: mp 222-224 °C; ¹H NMR (DMSO-d₆, 270 MHz) & 1.67-1.75 (m, 4 H), 2.03-2.26 (m, 2 H), 2.46-2.52 (m, 2 H), 2.64 (s, 2 H), 2.81 (t, 2 H, J = 6.9 Hz), 3.58 (m, 2 H), 4.49-4.57 (m, 1 H), 7.44 and 7.94 (AA'BB', 4 H), 8.67 (d, 1 H, J = 7.6 Hz), 9.77 (s, 1 H), 9.88 (t, 1 H, J = 5.5 Hz),10.40 (s, 1 H), 12.52 (br s, 2 H); HRFABMS calcd for C21H28N5O7 (MH^+) 460.1832, found 460.1834. Anal. Calcd for $C_{21}H_{25}N_5O_7$: C, 54.90; H, 5.48; N, 15.24. Found: C, 55.14; H, 5.40; N, 15.04.

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Supplementary Material Available: ¹H NMR spectra for compounds 4, 10, 11, 13-19, 22, 24-34, 38-45, 49, 54, and 57 and IR spectral data for compounds 4-6, 8-11, 13, 14, 16-19, 21, 22, 24-45, 47-49, 51, 53, 54, and 56-61 (42 pages). Ordering information is given on any current masthead page.

Triplex Formation of an Oligonucleotide Containing 2'-O-Methylpseudoisocytidine with a DNA Duplex at Neutral pH

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The synthesis of the hexadecanucleotide 5'TTTT1TTTT111111T3' (1-16mer) containing 2-amino-5-(2-Omethyl-β-D-ribofuranosyl)-4(1H)-pyrimidinone (2'-O-methylpseudoisocytidine or 1) is described. Triplex formation of 1-16mer with a deoxyribonucleotide duplex 5'd(ACCAAAAGAAAAGGGGGGGACCA)3'-5'd-(TGGTCCCCCCTTTTCCTTTTGGT)3' (duplex-22) which contains the "polypurine tract" found in the genome of human T-cell leukaemia (lymphotropic) virus (HTLV-III) was studied by thermal denaturation and circular dichroism (CD) spectra in aqueous solution at neutral pH. The "polypurine tract" contains a homoguanine cluster consisting of 6 deoxyguanine residues. The results indicate that 1-16mer and duplex-22 formed a triplex at neutral condition (0.01 M Na cacodylate, 0.5 M NaCl, 5 mM MgCl₂, pH 7.2. T_m = 20 °C). In contrast, a hexadecadeoxynucleotide, 5"TTTTMTTTTMMMMMMT3' (M-16mer), containing 5-methyl-2'-deoxycytidine (M) did not form a stable triplex with duplex-22 at the same condition ($T_m < 0$ °C). The CD mixing titration indicated that the triplex was formed with 1:1 (duplex:third strand) molecular stoichiometry.

Introduction

The formation of triple-stranded nucleic acid helices (triplex) has been studied for more than three decades.¹⁻³ In recent years, studies of sequence-specific triplex formation of short synthetic oligonucleotides (or their analogues) are of prime interest because the approach is applicable to biological and biochemical studies such as site specific cleavage of DNA,⁴⁻⁷ inhibition of DNA-protein binding,⁸⁻¹⁰ and inhibition of gene expression.¹¹

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The most common motif is that each case triad of a triplex consists of two pyrimidine residues from two homopyrimidine strands and a purine residue from a homopurine strand. The third strand (i.e., the second pyrimidine strand) is located in the major groove of a duplex consisting of Watson-Crick base pairing, and its sugarphosphate backbone polarity is parallel to that of the purine strand.^{4,12-16} Thymines or cytosines in the third

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